For reprint orders, please contact: reprints@future-science.com



A point-of-care global thrombosis test measuring occlusion time and endogenous lysis time may indicate thrombotic status

Junichiro Yamamoto^{*,1}, Nobutaka Inoue², Kazunori Otsui³, Hideo Ikarugi⁴, Muneshige Shimizu⁵, Shinji Yamamoto⁶, Masahiro Murakami⁷, Yoshinobu Ijiri⁸ & Kjell S Sakariassen⁹

¹Kobe Gakuin University, Kobe 651–2180, Japan

²Department of Cardiovascular Medicine, Kobe Rosai Hospital, Kobe 651–0053, Japan

³Department of General Internal Medicine, Kobe University Hospital, Kobe 650–0017, Japan

⁴School of Economics & Management, University of Hyogo, Kobe 651–2197, Japan

⁵Department of Fisheries, Faculty of Marine Science & Technology, Tokai University, Shizuoka 424–8610, Japan

⁶Faculty of Sport Sciences, Nihon Fukushi University, Aichi 470–3295, Japan

⁷Laboratory of Pharmaceutics, Faculty of Pharmacy, Osaka Ohtani University, Osaka 584–8540, Japan

⁸Department of Health & Nutrition, Faculty of Health & Nutrition, Osaka Shoin Women's University, Osaka 577–8550, Japan

⁹Str. Campo e Zampe 12, I–13900, Biella, BI, Italy

*Author for correspondence: yamamoto@nutr.kobegakuin.ac.jp

" implication of these models in the pharma industry has reduced costs of the discovery and developmental activities and the need for animal experiments."

First draft submitted: 29 April 2019; Accepted for publication: 23 May 2019; Published online: 2 July 2019

Keywords: blood flow • endogenous fibrinolysis • native blood • platelet aggregation • shear • thrombosis • thrombotic biomarker • thrombotic indicator

Can a single molecular biomarker predict overall thrombotic status?

Much research to identify clinically useful molecular biomarkers that reflect overall thrombotic status has been performed over the last few decades. One of the most probable molecular biomarkers appears to be D-dimer, but this lacks specificity, with poor predictive value in arterial thrombosis [1]. It is doubtful whether any single molecular biomarker could predict a multifactorial event such as an arterial thrombotic event. Another important problem is that *in vitro* anticoagulation of blood sample affects the results of the test used and renders it rather unphysiological [2–8].

However, some promising nonanticoagulated global hemostasis and thrombosis tests have been developed and have been used successfully in diagnosis and medical discovery and developments [9–14]. These tests have been well developed and proven to be highly important for both diagnostic and therapeutic purposes. Blood is drawn directly from an antecubital vein over a prothrombotic surface consisting of either human type III collagen fibrils or human tissue factor/phospholipids, which promotes thrombus formation by activation of both platelets and coagulation. Thrombus formation is performed at vessel wall shear rates varying from 100 to 32,000 s⁻¹. The impact of vessel wall shear rates on the various mechanisms of thrombus formation and on anticoagulant and antiplatelet agents are very high. Platelet activation is increased by increasing arterial wall shear rate, whereas coagulation is increased by low venous wall shear rates. These physical shear rate effects have important effects on the activities of many antiplatelet and anticoagulant agents. Also, impacts of cigarette smoking and physical activities on thrombus formation have been studied with this global nonanticoagulated blood test [15,16].

It should also be mentioned that the use of these human nonanticoagulated blood tests is of importance for the antithrombotic and hematologic pharma industry, since animal studies may not always predict the human reaction. Furthermore, implication of these models in the pharma industry has reduced costs of the discovery and developmental activities and the need for animal experiments.



Suggestions from animal experiments

We have assessed the optimal clinical test using helium neon laser-induced thrombosis *in vivo* tests and shearinduced platelet-rich thrombosis/thrombolysis (fibrinolysis) *in vitro* tests with nonanticoagulated (native) blood sample in experimental animals.

We screened antithrombotic activity of fresh juice prepared from 15 carrot varieties and further analyzed antithrombotic activity of the juice from the selected three varieties before and after heat treatment at 100° C for 10 min by the shear-induced platelet-rich thrombosis/fibrinolysis in vitro test (global thrombosis test [GTT]). Results for the first variety (SAKATA-0418) obtained by GTT in vitro test predicted that this variety has antithrombotic activity before heat treatment due to shortened lysis time (LT), but prothrombotic activity after heat treatment due to shortened occlusion time (OT). The prediction was proved by helium neon laser-induced thrombosis in vivo test. The result for the second variety (SAKATA-0420) before heat treatment as obtained by GTT could not predict pro- and antithrombotic activities because OT predicted prothrombotic activity, but LT predicted antithrombotic activity. This was demonstrated to be prothrombotic activity by helium neon laser-induced in vivo test, that is, platelet-rich thrombosis activity (OT) was predominant over thrombolytic activity (LT). Results after heat treatment predicted that overall activity may be antithrombotic because OT was not changed but LT was shortened. This was demonstrated by helium neon laser-induced thrombosis in vivo test. The result for the third variety (SAKATA-0421) before heat treatment predicted antithrombotic activity due to prolonged OT and shortened LT. This was demonstrated by helium neon laser-induced thrombosis *in vivo* test. Results after heat treatment predicted antithrombotic activity due to shortened LT regardless of unchanged OT. This was demonstrated by helium neon laser-induced thrombosis in vivo test [17].

Overall, it was shown that shear-induced platelet-rich thrombosis/fibrinolysis *in vitro* tests using native blood (nonanticoagulated blood) are well-matched to helium neon laser-induced platelet-rich thrombosis *in vivo* test [17]. In addition, the GTT using native blood and enabling simultaneous measurement of platelet-rich thrombotic and fibrinolytic activities demonstrated that thrombotic status is governed by both activities, indicating that simultaneous measurement of platelet-rich thrombosis and fibrinolysis measurement is useful for treatment of patients with thrombotic diseases. We have published articles on this topic in humans as well [6–8,18,19].

Comparison of thrombotic molecular biomarkers using anticoagulated blood sample & shear-induced thrombosis/fibrinolysis test using nonanticoagulated blood (native blood) sample

We compared molecular biomarkers, fibrinogen, Fibrinopeptide A (FPA), D-dimer and agonist-induced platelet aggregation tests using anticoagulated blood and a shear-induced thrombosis/fibrinolysis test (Thrombotic Status Analyser [TSA] Montrose Diagnostics Ltd, UK; shear stress: 400 dynes/cm²) using native blood samples in non-insulin-dependent diabetic and stroke patients [20].

The thrombotic status of diabetic patients classified into subgroups according to the criteria of nephropathy stages and stroke patients with hemorrhage, lacunar infarction or other complications was compared by biomarkers using anticoagulated blood and a shear-induced thrombosis/fibrinolysis test using native blood.

Collagen-induced whole blood platelet aggregation test showed lowered platelet reactivity in all subgroups compared with control, suggesting antithrombotic status. ADP-induced platelet aggregation test showed various results; prothrombotic or antithrombotic status or no difference compared with control. FPA measurement gave no informative results. D-dimer measurement is a sensitive test, but this test was not specific, therefore practically may not be useful for treatment of patients with thrombotic disorders. In addition, it is unclear that increase in D-dimer is due to enhanced fibrin formation or enhanced fibrin degradation, or both. This may make it difficult to treat patients with thrombotic disorders, that is, it is unclear whether treatment should be targeted toward fibrin formation or fibrinolysis. In addition, information on platelet reactivity, which plays a central role in arterial thrombosis, is lacking in D-dimer measurement tests.

In contrast, shear-induced thrombosis/fibrinolysis tests using native blood sample can clarify whether prothrombotic or antithrombotic status is due to platelet-rich thrombotic activity or fibrinolytic activity. GTT is sensitive and can show prothrombotic status after physiological stress such as over work. In contrast, the widely used tests PT and APTT could not show significant prothrombotic status [19].

A shear-induced platelet-rich thrombosis test using native blood was superior in detecting prothrombotic status compared with the agonist-induced platelet aggregation tests and molecular biomarkers. Neither FPA nor D-dimer proved to be a reliable biomarker of prothrombotic status. Extensive clinical research has been performed with GTT showing the clinical effectiveness of this test in identifying patients at high risk.

Conclusion

The point-of-care shear-induced thrombosis and fibrinolysis test using native blood samples (GTT) can measure platelet-rich thrombotic and endogenous fibrinolytic activities and thrombus stability under blood flow [21]. Unique features of the GTT include the testing of native blood and initiation of thrombus formation solely by pathologically relevant high shear stress. Formation of fibrin-stabilized thrombus and its lysis are detected – more information can be found in [7].

We propose that the thrombosis OT and endogenous LT measured simultaneously in the GTT are good clinical indicators to assess multifactorial events of thrombotic status. GTT is useful for not only treating patients with thrombotic disorders including cancer-associated thrombosis, but also in prevention of diseases and applications in other fields such as agriculture, nutrition, exercise and other aspects of healthcare.

Author contributions

J Yamamoto and KS Sakariassen were responsible for conception and design. Manuscript writing was undertaken by J Yamamoto and KS Sakariassen. All authors revised and approved the final version of the manuscript.

Financial & competing interests disclosure

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

No writing assistance was utilized in the production of this manuscript.

Open access

This work is licensed under the Creative Commons Attribution 4.0 License. To view a copy of this license, visit http://creativecomm ons.org/licenses/by/4.0/

References

- Donkel SJ, Benaddi B, Dippel DWJ, Ten Cate H, de Maat MPM. Prognostic hemostasis biomarkers in acute ischemic stroke. Arterioscler. Thromb. Vasc. Biol. 39(3), 360–372 (2019).
- 2. Paniccia R, Priora R, Liotta AA, Abbate R. Platelet function tests: a comparative review. Vasc. Health Risk Manag. 11, 133–148 (2015).
- 3. Saraf S, Christopoulos C, Salha IB, Stott DJ, Gorog DA. Impaired endogenous thrombolysis in acute coronary syndrome patients predicts cardiovascular death and nonfatal myocardial infarction. *J. Am. Coll. Cardiol.* 55(19), 2107–2115 (2010).
- Gorog DA, Fuster V. Platelet function tests in clinical cardiology: unfulfilled expectations. J. Am. Coll. Cardiol. 61(21), 2115–2129 (2013).
- Okafor ON, Gorog DA. Endogenous fibrinolysis: an important mediator of thrombus formation and cardiovascular risk. J. Am. Coll. Cardiol. 65(16), 1683–1699 (2015).
- 6. Yamamoto J, Tamura Y, Ijiri Y, Iwasaki M, Murakami M, Matsuo O. Evaluation of antithrombotic effect: importance of testing components and methodologies. *Drug Discov. Ther.* 9(4), 258–266 (2015).
- 7. Yamamoto J, Ijiri Y, Ikarugi H, Otsui K, Inoue N, Sakariassen KS. Prevention of thrombotic disorders by antithrombotic diet and exercise: evidence by using global thrombosis tests. *Future Sci. OA* 4(4), FSO285 (2018).
- Yamamoto J, Ijiri Y, Shimizu M, Yamamoto S, Ikarugi H, Sakariassen KS. Shear rate-dependent thrombosis/fibrinolysis tests using non-anticoagulated blood could be useful in the prevention of thrombotic disorders. *Future Sci. OA* 5(2), FSO357 (2018).
- 9. Sakariassen KS. Blood flow devices in medical research and clinical testing in humans: are we approaching personalized medicine? *Future Cradiol.* 3(1), 71–90 (2007).
- 10. Sakariassen KS, Turitto VT, Baumgartner HR. Recollection of the development of flow devices for studying mechanisms of hemostasis and thrombosis in flowing whole blood. J. Thromb. Haemost. 2(10), 1681–1690 (2004).
- 11. Sakariassen KS, Hanson SR, Cadroy Y. Methods and models to evaluate shear-dependent and surface reactivity-dependent anti-thrombotic efficacy. *Thromb. Res.* 104(3), 149–174 (2001).
- 12. Zwaginga JJ, Nash G, King MR *et al.* Flow-based assays for global assessment of hemostasis. Part 1. J. Thromb. Haemost. 4(11), 2486–2487 (2006).
- 13. Zwaginga JJ, Sakariassen KS, Nash G et al. Flow-based assays for global assessment of hemostasis. Part 2. J. Thromb. Haemost. 4(12), 2716–2717 (2006).

- 14. Sakariassen KS, Orning L, Turitto VT. The impact of blood shear rate on arterial thrombus formation. *Future Sci. OA* 1(4), FSO30 (2015).
- 15. Cadroy Y, Pillard F, Sakariassen KS, Thalamas C, Boneu B, Riviere D. Strenous but not moderate exercise increases the thrombotic tendency in healthy sedentary male volunteers. *J. Appl. Physiol.* 93(3), 829–833 (2002).
- Roald HE, Orvim U, Bakken IJ, Barstad RM, Kierulf P, Sakariassen KS. Modulation of thrombotic responses in moderately stenosed arteries by cigarette smoking and aspirin ingestion. *Arterioscler. Thromb.* 14(4), 617–621 (1994).
- 17. Yamamoto J, Naemura A, Ijiri Y *et al.* The antithrombotic effects of carrot filtrates in rats and mice. *Blood Coagul. Fibrinolysis* 19(8), 785–792 (2008).
- Taomoto K, Ohnishi H, Kuga Y et al. Platelet function and spontaneous thrombolytic activity of patients with cerebral infarction assessed by the global thrombosis test. Pathophysiol. Haemost. Thromb. 37(1), 43–48 (2010).
- Otsui K, Yamamoto J, Inoue N. Overwork accelerates thrombotic reaction: implications for the pathogenesis of Karoshi. J. Thromb. Thrombolysis 45(2), 222–224 (2018).
- 20. Nakajima S, Noguchi T, Taka T *et al.* A global platelet test of thrombosis and thrombolysis detects a prothrombotic state in some patients with non-insulin dependent diabetes and in some patients with stroke. *Platelets* 11(8), 459–466 (2000).
- 21. Spinthakis N, Farag M, Gue YX, Srinivasan M, Wellsted DM, Gorog DA. Effect of P2Y12 inhibitors on thrombus stability and endogenous fibrinolysis. *Thromb. Res.* 173, 102–108 (2019).